科技部補助專題研究計畫成果報告

期末報告

利伯氏遺傳視神經病變之雌激素相關受器與性別差異研究

計	畫	類	別	:	個別型計畫
計	畫	編	號	:	MOST 103-2629-B-075-001-
執	行	期	間	:	103年08月01日至104年07月31日
執	行	單	位	:	臺北榮民總醫院眼科部

計畫主持人: 王安國

計畫參與人員: 學士級-專任助理人員:林珮蕓

處理方式:

1. 公開資訊:本計畫涉及專利或其他智慧財產權,2年後可公開查詢

- 2.「本研究」是否已有嚴重損及公共利益之發現:否
- 3.「本報告」是否建議提供政府單位施政參考:否

中華民國104年10月16日

中 文 摘 要 : 利伯氏遺傳視神經病變(LHON)是一種母系遺傳的粒線體突變,它會 造成急性視力減退,好發於男性,佔80%,視力常惡化至零點一以下 ,伴隨視野缺損,兩眼可同時發病。粒線體突變會傳給所有母系細 胞,然而具有突變基因的男性發病率僅有50%,女性更只有10%,顯 示其發病率低與好發於男性的特點。

其發病率低與好發於男性的特點,目前無明確原因,推測可能 源於神經保護機轉,若能瞭解此機轉可能有助於未來的治療,雌激 素與其衍生物已証明具有神經保護作用,我們提出此計畫來探討雌 激素相關受器(Estrogen-related receptor, ERRs)在利伯氏遺傳視 神經病變致病機轉與其發病率性別差異所扮演的角色,特別著重於 神經保護與粒線體生物生成(mitochondria biogenesis)功能。我們 利用螢光免疫染色,來檢查三種雌激素相關受器(ERR α , ERR β and ERR γ)在小鼠視網膜的表現,發現ERR α 和ERR γ 蛋白在小鼠視網膜 全層都有表現,特別在節細胞層(ganglion cell layer)與感光細胞 內層(inner segment of photoreceptor)有較強表現,視網膜節細 胞(retinal ganglion cell)與無軸細胞(amacrine cell)都會表現 不等程度的ERR α 和ERR γ 蛋白。此外,ERR β 主要表現在從神經纖維 層(nerve fiber layer)延伸到外叢層(outer plexiform layer)的 視網膜內層,而其外的外層視網膜沒有表現此蛋白,ERR β 的表現與 Muller細胞纖維範圍相符,顯示此蛋白主要表現在Muller細胞。

我們施行視神經壓迫傷手術(optic nerve crush),檢查小鼠視 網膜在受傷後之三種雌激素相關受器表現變化,發現三種雌激素相 關受器表現有類似的變化趨勢,在受傷後,它們在節細胞層都逐漸 增加表現量,第三天達到最高量,隨後漸漸降低,在第五到七天會 低於對照組。然而,此三種雌激素相關受器的表現變化,在雄鼠與 雌鼠的表現相似,並無性別差異。最後,我們採用西方墨點試驗 ,檢驗雌激素相關受器的表現變化,發現其變化趨勢與螢光免疫染 色相同,這樣的變化趨勢,代表神經細胞對於軸突損傷所產生的細 胞反應。

中文關鍵詞:利伯氏遺傳視神經病變,雌激素相關受器

英 文 摘 要: Leber's hereditary optic neuropathy (LHON) is a maternally transmitted disease caused by mitochondria DNA (mtDNA) mutation. It is characterized by acute and subacute visual loss predominantly affecting young man. Vision usually deteriorates to the degree of less than 20/200, accompanied with a cecocentral scotoma. Both eyes are involved with or without intervals. The mtDNA mutation is transmitted to all the maternal lineages. However, only approximately 50% of men and 10% of women harboring a pathogenic mtDNA mutation develop optic neuropathy, reflecting both the incomplete penetrance and its unexplained male prevalence that over 80% of patients are male.

The incomplete penetrance and male prevalence is still the major unexplained issue in LHON. These low penetrance and male prevalence may result from undefined neuroprotective mechanism which may be beneficial for

future treatment. Estrogen and its non-feminizing analogues have already proved its efficacy in the neuroprotection. We propose to investigate the role of estrogen and esotrogenrelated receptor (ERRs) family in the gender difference of LHON and its potential role in neuroprotection and mitochondrial biogenesis. We have examined the expression of estrogen-related receptors (ERR α , ERR β and ERR γ) in mouse retina. ERR α and ERR γ proteins are both widely expressed over the whole retinal layers, with intense level over ganglion cell layer and photoreceptor inner segment. Both retinal ganglion cell and amacrine cell may express various amounts of ERR α and ERR γ proteins. ERR β is expressed mainly over inner retina extending from nerve fiber layer to the outer plexiform layer, with the outer retina relatively spared. Its expression corresponds to the span of Muller cell processes.

Following optic nerve injury, we observe the expressional change of estrogen-related receptors (ERR α , ERR β and ERR γ) in the adult mice retina in response to optic axonal injury. All three estrogen-related receptors showed a similar trend that their expression in the ganglion cell layer increased gradually and peaked around 3 days after injury, and then declined at 5 days to 7 days thereafter. Nevertheless, we did not observe any gender difference of estrogen-related receptors expression throughout the whole course. Western blot analysis showed a similar pattern of expressional change after optic nerve injury, which may reflect neuronal cell response to axonal injury.

英文關鍵詞:Leber's hereditary optic neuropathy, estrogen-related receptor

科技部補助專題研究計畫成果報告

(□期中進度報告/Ⅳ期末報告)

利伯氏遺傳視神經病變之雌激素相關受器與性別差異研究

計畫類別: ♥個別型計畫 □整合型計畫 計畫編號: MOST 103-2629-B-075-001-執行期間: 103 年 8 月 1 日至 104 年 7 月 31 日

執行機構及系所:台北榮民總醫院眼科部

計畫主持人:王安國

本計畫除繳交成果報告外,另含下列出國報告,共_0_份: □執行國際合作與移地研究心得報告

□出席國際學術會議心得報告

期末報告處理方式:

1. 公開方式:

□非列管計畫亦不具下列情形,立即公開查詢

□涉及專利或其他智慧財產權,□一年 Ⅴ二年後可公開查詢

2.「本研究」是否已有嚴重損及公共利益之發現:▼否□是

3.「本報告」是否建議提供政府單位施政參考 Ⅳ否 □是, (請列舉提 供之單位;本部不經審議,依勾選逕予轉送)

中華民國 104 年 10 月 16 日

1

目錄

I.	中文摘要	р.3
II.	英文摘要	p.4
III	[. 報告內容	.p.5
IV	7. 參考文獻	.p.19
V.	成果自評	.p.24

一.中文摘要

利伯氏遺傳視神經病變(LHON)是一種母系遺傳的粒線體突變,它會造成急性視力 減退,好發於男性,佔80%,視力常惡化至零點一以下,伴隨視野缺損,兩眼可同時發 病。粒線體突變會傳給所有母系細胞,然而具有突變基因的男性發病率僅有50%,女性 更只有10%,顯示其發病率低與好發於男性的特點。

其發病率低與好發於男性的特點,目前無明確原因,推測可能源於神經保護機轉, 若能瞭解此機轉可能有助於未來的治療,雌激素與其衍生物已証明具有神經保護作用, 我們提出此計畫來探討雌激素相關受器(Estrogen-related receptor, ERRs)在利伯氏遺傳 視神經病變致病機轉與其發病率性別差異所扮演的角色,特別著重於神經保護與粒線體 生物生成(mitochondria biogenesis)功能。我們利用螢光免疫染色,來檢查三種雌激素相 關受器(ERRα, ERRβ and ERRγ)在小鼠視網膜的表現,發現ERRα和ERRγ蛋白在小鼠視 網膜全層都有表現,特別在節細胞層(ganglion cell layer)與感光細胞內層(inner segment of photoreceptor)有較強表現,視網膜節細胞(retinal ganglion cell)與無軸細胞(amacrine cell)都會表現不等程度的ERRα和ERRγ蛋白。此外,ERRβ主要表現在從神經纖維層 (nerve fiber layer)延伸到外叢層(outer plexiform layer)的視網膜內層,而其外的外層視 網膜沒有表現此蛋白,ERRβ的表現與Muller細胞纖維範圍相符,顯示此蛋白主要表現 在Muller細胞。

我們施行視神經壓迫傷手術(optic nerve crush),檢查小鼠視網膜在受傷後之三種雌 激素相關受器表現變化,發現三種雌激素相關受器表現有類似的變化趨勢,在受傷後, 它們在節細胞層都逐漸增加表現量,第三天達到最高量,隨後漸漸降低,在第五到七天 會低於對照組。然而,此三種雌激素相關受器的表現變化,在雄鼠與雌鼠的表現相似, 並無性別差異。最後,我們採用西方墨點試驗,檢驗雌激素相關受器的表現變化,發現 其變化趨勢與螢光免疫染色相同,這樣的變化趨勢,代表神經細胞對於軸突損傷所產生 的細胞反應。

二. 英文摘要

Summary

Leber's hereditary optic neuropathy (LHON) is a maternally transmitted disease caused by mitochondria DNA (mtDNA) mutation. It is characterized by acute and subacute visual loss predominantly affecting young man. Vision usually deteriorates to the degree of less than 20/200, accompanied with a cecocentral scotoma. Both eyes are involved with or without intervals. The mtDNA mutation is transmitted to all the maternal lineages. However, only approximately 50% of men and 10% of women harboring a pathogenic mtDNA mutation develop optic neuropathy, reflecting both the incomplete penetrance and its unexplained male prevalence that over 80% of patients are male.

The incomplete penetrance and male prevalence is still the major unexplained issue in LHON. These low penetrance and male prevalence may result from undefined neuroprotective mechanism which may be beneficial for future treatment. Estrogen and its non-feminizing analogues have already proved its efficacy in the neuroprotection. We propose to investigate the role of estrogen and esotrogen-related receptor (ERRs) family in the gender difference of LHON and its potential role in neuroprotection and mitochondrial biogenesis. We have examined the expression of estrogen-related receptors (ERR α , ERR β and ERR γ) in mouse retina. ERR α and ERR γ proteins are both widely expressed over the whole retinal layers, with intense level over ganglion cell layer and photoreceptor inner segment. Both retinal ganglion cell and amacrine cell may express various amounts of ERR α and ERR γ proteins. ERR β is expressed mainly over inner retina extending from nerve fiber layer to the outer plexiform layer, with the outer retina relatively spared. Its expression corresponds to the span of Muller cell processes.

Following optic nerve injury, we observe the expressional change of estrogen-related receptors (ERR α , ERR β and ERR γ) in the adult mice retina in response to optic axonal injury. All three estrogen-related receptors showed a similar trend that their expression in the ganglion cell layer increased gradually and peaked around 3 days after injury, and then declined at 5 days to 7 days thereafter. Nevertheless, we did not observe any gender difference of estrogen-related receptors expression throughout the whole course. Western blot analysis showed a similar pattern of expressional change after optic nerve injury, which may reflect neuronal cell response to axonal injury.

三. 報告內容

3.1 緣由與目的

A.Background

Leber's hereditary optic neuropathy (LHON)¹

LHON is a maternally transmitted disease characterized by acute and subacute visual loss predominantly affecting young man.¹⁻³ It usually onsets between 15 to 35 years of age, with a male predominance.^{1,4} The course of visual loss is usually acute or subacute. The optic disc became hyperemic and associated with peripapillary telangiectasia. The retinal nerve fiber layers are swollen. Over months, the disc edema subsided and became pallor and atrophic. Vision deteriorated to the degree of less than 20/200, commonly accompanied with a cecocentral scotoma.¹ Both eyes are involved with or without intervals.¹

Three primary mitochondrial DNA (mtDNA) mutations underlie the main pathogenesis of LHON. The first association of a mtDNA 11778 mutation with LHON was reported by Wallace and colleagues in 1988.⁵ These three primary mutation of mtDNA 11778, 14484 and 3460 encodes the NADH dehydrogenase subunit 4, subunit 6 and subunit 1 of Complex I of the respiratory chain respectively.⁵⁻⁸

Male prevalence of LHON¹

Despite intense studies on the clinical and molecular aspects of LHON from 1988, the pathogenesis is still unclear especially in the area of gender prevalence and penetrance.¹ The mtDNA mutation is transmitted to all the maternal lineages. However, only approximately 50% of men and 10% of women harboring a pathogenic mtDNA mutation develop optic neuropathy, reflecting both the incomplete penetrance and the gender prevalence difference.¹ LHON is well known for its male prevalence that over 80% of patients are male.^{1,2} This male predominance exists in different mutation groups with a male to female ratio of 3:1, 4–6:1 and 8:1 in patients harboring 3460, 11778 and 14484 mutations, respectively.^{1,4} The reason for this male predominance remains unknown.

Penetrance of LHON¹

The penetrance in LHON is incomplete and variable that a positive family history was found in 50% of patients with 11778 mutation, 71% with 3460 mutation and 100% with 14484 mutation.^{1,9} The penetrance of LHON is variable even with the same mutation in homoplasmic fashion within the same family in different pedigree branch.^{1,10} All these features can not be explained by a single point mutation of mtDNA alone. Thus, genetic and

epigenetic factors have been presumed to be involved in the penetrance of the LHON. Prior investigated genetic modifiers include heteroplasmy,¹¹⁻¹⁴ secondary mtDNA mutations,¹⁵⁻¹⁶ mtDNA haplogroup,¹⁷⁻¹⁹ X-linked modifying gene or susceptibility locus,²⁰⁻²² and other nuclear genes.²³ Tobacco and alcohol consumption were considered as epigenetic factors which may affect the penetrance as well.²⁴⁻²⁵

Estrogen and its receptors

Estrogens are a family of cholesterol-derived steroid hormones that may regulate growth and differentiation in various tissues including reproductive system and central nervous system.²⁶⁻²⁸ Its action was mediated through estrogen receptors (ERs). Estrogens enter the nucleus of the target cell and then fuse with ERs to make an estrogen-ER complex, which in turn may react with the estrogen response element (ERE) of the target genes. Then the transcription of these estrogen-dependent genes will be activated.

Two major estrogen receptors, ERalpha and ERbeta, exist on various tissues. ERalpha, encoded by ESR1 gene on chromosome 6q25, is the major receptor in the adult uterus and responsible for the proliferative reaction in uterus.^{29,30} The third estrogen receptor, GPR30, has been found recently with its exact function remaining to be elucidated.^{31,32}

Estrogen and Neuroprotection

Estrogen has been shown to have neurotrophic and neuroprotective effect in various tissues.³³ In retinal pigment epithelium cells, 17-beta estradiol protects the cells from oxidative stress through the ERbeta-dependent mechanism. The cytoprotection of estrogen for retinal neurons occurs by reduction of ROS production, induction of cellular antioxidant genes, and preservation of mitochondrial function.³⁴⁻³⁷

Giordano C. et al. tried to explore the reasons for the higher prevalence of LHON in males and they proposed the potential compensatory effect of estrogen on mutant cell metabolism may underlie the gender prevalence.³⁸ They have provided a possible metabolic basis for the unexplained male predominance in LHON. It indicates that estrogen do have a neuroprotective effect on LHON. However, the exact mechanism of how the estrogen influence gender difference in LHON remains unknown.

Estrogen-Related Receptor alpha (ERRalpha)³⁹

The esotrogen-related receptor (ERR) family is consisted of three major family members (alpha, beta, and gamma).³⁹⁻⁴¹ They possess the typical structure of nuclear receptors, which contains a zinc finger DNA binding domain and a conserved C-terminal domain with a putative ligand binding domain and interaction surfaces for coactivators and corepressors.³⁹ These ERR members have a significant homology in DNA binding domain to the ERalpha, and with lesser similarity of the N- and C- terminal.⁴² Estrogen has also been shown to stimulate the ERRalpha gene expression, ⁴² which was found to bind many

different ER response elements (EREs).⁴³⁻⁴⁵ In addition, it could interact with similar coactivators of ERalpha.^{44,45} Thus, ERRalpha gene may act as a downstream target of ERalpha.⁴²

ERRalpha and Neuroprotection

PGC-1alpha (Peroxisome proliferator activated receptor Gamma Coactivator 1-alpha) is a master regulator for many cellular functions including mitochondrial biogenesis, metabolism, suppression of ROS, and stress response.⁴⁶⁻⁴⁸ PGC-1alpha may promote the SIRT3 expression through ERRalpha receptor, which binds to and induces SIRT3 promoter.⁴⁹ Up-regulated SIRT3 may in turn deacetylate and activate mitochondrial enzymes involved in fatty acid beta-oxidation, amino acid metabolism, electron transport chain, and antioxidant defenses.⁴⁸ On the other hand, SIRT3 may also up-regulate the expression of PGC-1alpha by a positive feedback system mediated by AMP-activated kinase.⁵⁰⁻⁵² In addition to the increase of mitochondrial energy metabolism, SIRT3 prevents apoptosis by lowering reactive oxygen species and decreasing the formation of mitochondrial permeability transition pore.⁴⁸ Both the antioxidant effect and the anti-apoptotic effect mediated by SIRT3 suggest a neuroprotective role of ERRalpha which may improve neuronal survival and reduce aging effect in cell.^{48,53}

The pathogenesis of male prevalence in LHON is still unclear yet.¹ We hypothesize that estrogen may play a role in this gender difference through neuroprotective function with unclear mechanism. In the current project, we propose to investigate the role of ERRs, a possible downstream target of estrogen, in the pathogenesis of gender difference in LHON. In addition, we will also investigate its potential role in neuroprotection and mitochondrial biogenesis.

3.2 研究方法舆過程

Material and Methods

Animal and surgery

Adult Balb/c mice will be kept in the animal facility of our hospital. Institute guideline will be followed on handling of animals. Mice will be given food and water ad libitum. Adult mice that are at least two months old and body weight over 25 g, will be intraperitonially anesthetized with a combination of Zoletil 50 (0.1 mg/ 20 g body weight) and Rompun (0.023 mg/20 g body weight). Surgery is done under sterile conditions using a stereomicroscope. A conjunctival incision is made over the dorsal aspect of one eye, which is then gently rotated downward in the orbit. The orbital muscles are teased and deflected aside

to expose the optic nerve at its exit from the globe, which is then crushed twice with jewelers forceps near the back of the eye (within 0.5 mm). Care is taken not to damage the ophthalmic artery and °C retrobulbar sinus. The eye is then rotated back into position and rinsed with sterile saline and covered with 0.3% garamycin ointment. The mice are kept warm in cage till recovery. Mice are sacrificed at different time points (day 1, day 3, day 5 and day 7) using cervical dislocation under anesthesia.

Immunohistochemistry

(a) **Preparation and Fixation**

Adult mice eyes were enucleated and maintained in Hank's solution. Posterior eyecups were fixed with 4% paraformadehyde in 0.1M phosphate buffer (pH 7.25) for 1 hr at 4°C. Incubate the eyeballs with 30% sucrose/PBS overnight, then embed in OCT, and stored at -80° C till use. Sections were cut at 12 µm thick and collected on SuperFrost slides (Fisher Scientific, PA).

(b) Immunostaining

Sections are fixed with 2% paraformaldeyhyde for 20 min, and then pretreated with 0.3 % H_2O_2 in PBS for 15 min at room temperature. They are then incubated with 0.2% Trypsin in PBS for 5 min, and then with 0.3 % Triton X-100 in PBS for 5 min at room temperature. Sections are blocked with 5% BSA in PBS with 0.3 % Triton X-100 for 60 min at room temperature. Sections are incubated in primary antibodies, diluted in 5% BSA/PBS, for overnight at 4°C. They are then incubated with matched secondary antibodies for 2 hours at room temperature. Each step is preceded by washes in 1xPBS. Sections are then incubated with 1xPBS for three times. The sections are mounted with antifade medium (Citifluoro, Canada) and stored at 4°C.

For estrogen-related receptors, primary antibodies include: anti-ERR α antibody (Santa Cruz), anti-ERR β antibody (abcam), anti-ERR γ ntibody (Santa Cruz).

For retinal-specific markers, Anti-Brn3 (Santa Cruz), Anti-Brn3a (millipore), Anti-CHAT (abcam), Anti-GFAP (millipore), Anti-Glutamine synthetase (GS) (millipore).

For western blot internal control, anti-GAPDH (Santa Cruz).

(c) Image acquisition

Sections and coverslips will be analyzed with a Nikon Diaphot inverted microscope (E300, Nikon, Tokyo, Japan) equipped with a MicroMax cool CCD (Princeton Instrument, Trenton, NJ, USA). Image acquisition is performed using MetaMorph software (Universal Image

Corporation, Downingtown, PA, USA) with individual filter sets for each channel and are assembled with Adobe PhotoShop (Adobe Systems, CA).

Western Blot

Tissue samples will be loaded with an equal volume of 2X loading buffer (125 μ M Tris HCl, pH 6.8; 4% SDS; 20% glycerol and 10% 2-mercaptoethanol), boiled for 5 min and resolved by SDS-polyacrylamide gel (10%)electrophoresis.

Gels will be electro-blotted onto PVDF membranes (Millipore) in 25 mM Tris HCl, pH 8.3 containing 192 mM glycine, 20% methanol. Membranes will be blocked in PBS/0.1% tween (PBST) containing 5% fat-free dry milk (Carnation) for 1 hr. Primary antibody of anti-ERRalpha antibody, anti-ERRbeta antibody and anti-ERRgamma antibody will be incubated overnight in PBST with 5% dry milk. with Anti-GAPDH as internal control. After washing, HRP-conjugated secondary antibody will be incubated for 1 hr. Immunoreactive bands will be detected by enhanced chemiluminescence (ECL, Amersham) followed by exposure to Hyperfilm MP (Amersham).

3.3 研究結果

A. ERR Expression in Normal Mouse Retina

To verify the localization of estrogen-related receptors in the normal mouse retina, immunostaing was performed using anti- estrogen-related receptors (ERR α , ERR β and ERR γ) antibody. The result showed that ERR α protein is expressed on the retinal ganglion cell layer, inner plexiform layer, inner nuclear layer, outer plexiform layer and inner segment of photoreceptor (Fig 1A). ERR β protein is expressed on the nerve fiber layer, retinal ganglion cell layer, inner plexiform layer, inner nuclear layer and outer plexiform layer (Fig 1D). ERR γ protein is expressed on the nerve fiber layer, retinal ganglion cell layer, inner plexiform layer, inner nuclear layer, outer plexiform layer and inner segment of photoreceptor (Fig 1G). Both ERR α and ERR γ proteins are expressed over the whole retina, with high level of expression over ganglion cell layer and inner segment of photoreceptor. On the other hand, ERR β is expressed over inner retina extending from nerve fiber layer to the outer plexiform layer, which corresponds to the profile of Muller cell processes.

To further investigate if retinal ganglion cells (RGCs) express estrogen-related receptors (ERR α , ERR β and ERR γ), we have labeled adult mice retina with anti-Brn3 antibody, which is immune marker for RGCs. The results showed that ERR α and ERR γ proteins are expressed on both Brn3-positve and Brn3-negative cells in the ganglion cell layer. It implies that both RGCs and amacrine cells may express ERR α and ERR γ proteins. Meanwhile, the expressional level of ERR α and ERR γ proteins vary within these Brn3-positve cells. In addition, not all Brn3-postive cells express ERR α and ERR γ protein (Fig 1, C-C1, I-I1). ERR β protein is uniquely expressed on slender cell processes in the inner retina, extending around retinal ganglion cells (Fig 1, F-F1).



Figure 1. ERRs expression in adult mouse retina. Normal adult mouse were immunostained with anti-ERRα antibody (green, A-C), ant-ERRβ antibody (green, D-F) and anti-ERRγ antibody (green, G-I). Retinal ganglion cells are marked with Brn3 (red). DAPI (blue) was used as a nucleus marker. GCL: ganglion cell layer; INL: inner nuclear layer; ONL: outer nuclear layer.

To characterize displaced amacrine cells in the ganglion cell layer, we have labeled adult mice retina with anti-ChAT antibody, which is immune marker for amacrine cells. It is expressed on the amacrine cells of the inner nuclear layer and also the displaced amacrine cells in the ganglion cell layer (Fig 2, B,E,H). These amacrine cells exist on both sides of inner plexiform layer, also called as starburst amacrine cells. In addition, ChAT protein is also expressed as parallel lines in the inner plexiform layer, which implies the synapse connecting retinal ganglion cell and bipolar cells (Fig 2).

The results showed that ERR α and ERR γ proteins are expressed on both ChAT-positve and ChAT-negative cells in the ganglion cell layer. It implies that both amacrine cells and RGCs may express ERR α and ERR γ proteins. Meanwhile, ERR α and ERR γ proteins are expressed with different levels in these ChAT-positve cells, with some expressing high level and others none (Fig 2, C-C1, I-II). ERR β protein seems to be not expressed on the ChAT-positive cells (Fig 2, F-F1).



Figure 2. ERRs expression in adult mouse retina. Normal adult mouse were immunostained with anti-ERR α antibody (green, A-C), ant-ERR β antibody (green, D-F) and anti-ERR γ antibody (green, G-I). Starburst amacrine cells are marked with anti-ChAT antibody (red). DAPI (blue) was used as a nucleus marker. GCL: ganglion cell layer; INL: inner nuclear layer; ONL: outer nuclear layer.

From the above data, we learn that ERR β protein is present in the nerve fiber layer, retinal ganglion cell layer, inner plexiform layer, inner nuclear layer and outer plexiform layer. It is uniquely expressed over inner retina extending from nerve fiber layer to the outer plexiform layer, which corresponds to the profile of Muller cell processes. This specific expressional pattern indicates ERR β protein may be expressed exclusively in the retinal Muller cells. We have used two immune markers, GFAP and glutamine synthetase (GS), for double staining technique. GFAP is a general glial cell marker, while glutamine synthetase is a specific marker for Muller cells.

The results showed that in the ganglion cell layer, some ERR β -positive cells are GFAP immune-reactive. These GFAP-positive retinal astrocytes exist in the inner part of ganglion cell layer, with their GFAP-positive cell processes extending horizontally over the nerve fiber layer (Fig 3, B,E). ERR β protein is co-localized with GFAP protein on these retinal astrocytes, but not on the Muller cells. To further investigate, we co-stained glutamine synthetase (GS) with ERR β protein. The results showed that ERR β -positive cells are glutamine synthetase immune-reactive (Fig 3). Thus, Muller cell also express ERR β protein on their cell processes.



Figure 3. ERRβ expression in the retinal glia cells. Normal adult mouse were immunostained with ant-ERRβ antibody (green), GFAP (red, B,E) and anti-glutamine synthetase (red, H,K). GFAP is marker fro glia cell. Glutamine synthetase is marker for Muller cells. DAPI (blue) is used as a nucleus marker. GCL: ganglion cell layer; INL: inner nuclear layer; ONL: outer nuclear layer; IPL: inner plexiform layer.

B. Gender Effect on ERRs Expressional Change after Optic Nerve Crush

In this project, we try to investigate the expressional change of estrogen-related receptors (ERR α , ERR β and ERR γ) in a stressed retina. We plan to conduct optic nerve crush injury in adult male and female mouse and examine the gender difference of expressional change with immunohistochemistry. The immunostaining was performed to measure the amount of estrogen-related receptors expression in the retinal ganglion cell layer. The results showed that the expression of ERR α , ERR β and ERR γ increased in retinal ganglion cell layer at 3 days after optic nerve crush when compared with control group. However, the expression of ERR α , ERR β , ERR γ and Brn3 (RGC marker) decreased gradually thereafter at 5 days and

7 days after crush. But, there is no significant gender difference of ERR α , ERR β , ERR γ expression between male and female mice from 1 day to 7 days after optic nerve crush (Fig 4-6).



Figure 4. Gender Effect on ERR α **Expressional Change after Optic Nerve Crush** Optic nerve crush injury was performed in left eye of adult mice, with right eye spared as control. Both control and injured retina were labeled with anti-ERR α antibody (green). There is no significant difference between the crushed and control retina at 1 day after optic nerve injury (A1-D1). The expression of ERR α increased significantly in the crushed retina at 3 days post crush, as compared with control (A2-D2). The pattern of increased expression was similar among male and female mice. It decreased gradually in the crushed retina thereafter. ERR α expression in crushed retina became less as compared with control retina at 5 days and 7 days postcrush, with no gender difference (A3-D4).



Figure 5. Gender Effect on ERR β Expressional Change after Optic Nerve Crush Optic nerve crush injury was performed in left eye of adult mice, with right eye spared as control. Both control and injured retina were labeled with anti-ERR β antibody (green). There is no significant difference between the crushed and control retina at 1 day after optic nerve injury (A1-D1). The expression of ERR β increased significantly in the crushed retina at 3 days post crush, as compared with control (A2-D2). The pattern of increased expression was similar among male and female mice. It decreased gradually in the crushed retina thereafter. ERR β expression in crushed retina became less as compared with control retina at 5 days and 7 days postcrush, with no gender difference (A3-D4).



Figure 6. Gender Effect on ERRy Expressional Change after Optic Nerve Crush Optic nerve crush injury was performed in left eye of adult mice, with right eye spared as control. Both control and injured retina were labeled with anti-ERRy antibody (green). There is no significant difference between the crushed and control retina at 1 day after optic nerve injury (A1-D1). The expression of ERRy increased significantly in the crushed retina at 3 days post crush, as compared with control (A2-D2). The pattern of increased expression was similar among male and female mice. It decreased gradually in the crushed retina thereafter. ERRy expression in crushed retina became less as compared with control retina at 5 days and 7 days postcrush, with no gender difference (A3-D4).

C. ERRs Expressional Change after Optic Nerve Crush by Western Blot

We try to investigate the expression amount of estrogen-related receptors (ERR α , ERR β and ERR γ) by western blot. Retinal estrogen-related receptors (ERR α , ERR β and ERR γ) proteins show an increase at 3 days after optic nerve crush, and decrease at 5 days or 7 days after optic nerve crush, as compared with control retina. Interestingly, western blot analysis has a similar pattern of temporal change as those of immunostaining result (Fig 7).



Fig 7. Western Blot of estrogen-related receptors (ERR α , ERR β and ERR γ) Western blot analysis of retinal protein was performed with GAPDH as internal control. Crushed retina has an increased amount of ERRs at 3 days after crush, which decreased at 5 day and 7 days in comparison with control retina.

3.4 研究討論

LHON is the major hereditary optic neuropathy in Taiwan.¹ It has a minimum point prevalence for mtDNA LHON mutation of 11.82 per 100,000 subjects and the minimum point prevalence of visual failure due to LHON of 3.22 per 100,000 subjects in adults under 65 years of age at north-east England.^{1,54} It may cause bilateral blindness in a young adult and cause severe disability. Thus, it is of utmost importance to understand this disease. Though it is not difficult to diagnose since the development of molecular diagnosis, there has been few treatment available for this disease.

The incomplete penetrance and male prevalence is still the major unexplained issue in LHON. We propose this project to explore the role of estrogen-related receptors (ERR α , ERR β and ERR γ) in the pathogenesis of LHON, not only for the scientific interest but also for the possible therapeutic chance. The low penetrance in female LHON 11778 patients may results from some protective mechanism of estrogen and ERRs. If the underlying mechanism is well investigated, it could also benefit the future treatment. Estrogen and its non-feminizing analogues have already proved its efficacy in the neuroprotection. We expect there will be even more therapeutic options in the future if the mechanism for LHON gender difference is well understood.

We have examined the expression of estrogen-related receptors (ERR α , ERR β and ERR γ) in mouse retina. Expression of ERR α protein is found in the retinal ganglion cell layer, inner plexiform layer, inner nuclear layer, outer plexiform layer and inner segment of photoreceptor. ERR β protein is mainly expressed in the inner two third of mice retina, over the nerve fiber layer, retinal ganglion cell layer, inner plexiform layer, inner nuclear layer and outer plexiform layer. ERR γ protein expression is distributed over the nerve fiber layer, retinal ganglion cell layer, inner plexiform layer, outer plexiform layer, and error fiber layer, retinal ganglion cell layer, inner plexiform layer, outer plexiform layer and inner segment of photoreceptor. The expression patterns of ERR α and ERR γ proteins are similar that they are widely expressed over the whole retinal layers, with intense level over ganglion cell layer and photoreceptor inner segment. On the other hand, ERR β is expressed over inner retina with outer retina spared. It extends from nerve fiber layer to the outer plexiform layer, corresponding to the Muller cell processes. Double staining has shown that ERR β expression is co-localized with Muller cell marker glutamine synthetase.

With anti-Brn3 antibody (marker for RGCs) and anti-ChAT antibody (marker for amacrine cell), we have found that both RGCs and amacrine cells may express ERR α and ERR γ proteins. Meanwhile, the expressional level of ERR α and ERR γ proteins vary widely among these Brn3-positive or ChAT-positive cells. On the other hand, ERR β protein seems to be not expressed on the Brn3-positive or ChAT-positive cells

We try to investigate the expressional change of estrogen-related receptors (ERR α , ERR β and ERR γ) in a stressed retina after optic nerve crush. Gender difference in the expressional change of estrogen-related receptors was assessed as well. We found the expression of three estrogen-related receptors show a similar trend that its expression in the ganglion cell layer increased gradually and peaked at 3 days after optic nerve crush, and then decreased at 5 days and 7 days after crush injury. Nevertheless, we did not observe gender difference of estrogen-related receptors expression throughout the whole course. Western blot analysis showed a similar pattern of expressional change after optic nerve injury, which may reflect neuronal cell response to axonal injury.

四. 參考文獻 (References)

D. 參考文獻:

1. Yen MY, Wang AG, Wei YH. Leber's hereditary optic neuropathy: a multifactorial disease. Prog Retin Eye Res. 2006;25(4):381-96.

2.Newman NJ, Lott MT, Wallace DC. The clinical characteristics of pedigrees of Leber's hereditary optic neuropathy with 11778 mutation. Am J Ophthalmol 1991;111:750-762.

3.Newman NJ. Leber's hereditary optic neuropathy. New genetic consideration. Arch Neurol 1993;50:540-548.

4.Carelli V, Ross-Cisneros FN, Sadun AA. Mitochondrial dysfunction as a cause of optic neuropathies. Prog. Retin Eye Res 2004;23:53-89.

5.Wallace DC, Singh G, Lott MT, el al. Mitochondrial DNA mutation associated with Leber's hereditary optic neuropathy. Science 1988;242:1427-1430.

6.Huoponen K, Vilkki J, Aula P, Nikoskelainen EK, Savontaus M-L. A new mtDNA mutation associated with Leber hereditary optic neuropathy. Am J Hum Genet 1991;48:1147-1153.

7.Howell N, Bindoff A, McCullough DA, Kubacka I, Poulton J, Mackey D, Taylor L, Turnbull DM. Leber hereditary optic neuropathy: identification of the same mitochondrial ND1 mutation in six pedigrees. Am J Hum Genet 1991;49:939-950.

8.Mackey D, Howell N. A variant of Leber hereditary optic neuropathy characterized by recovery of vision and by an unusual mitochondrial genetic etiology. Am J Hum Genet 1992;51:1218-1228.

9.Riordan-Eva P, Sander MD, Govan GG, Sweeney MG, Costa JD, Harding AE. The clinical features of Leber's hereditary optic neuropathy defined by the presence of a pathogenic mitochondrial DNA mutation. Brain 1995;118:319-337.

10.Chalmers RM, Harding AE. A case-control study of Leber's hereditary optic neuropathy. Brain 119;1481-1486.

11.Smith KH, John DR, Heher KL, Miller NR. Heteroplasmy in Leber's hereditary optic neuropathy. Arch Ophthalmol 1993;111:1486-1490.

12.Jacobi FK, Leo-Kottler B, Mittelviefbaus K, Zrenner E, Myer J, Puscb CM, Wissinger B. Segregation patterns and heteroplasmy prevalence in Leber's hereditary optic neuropathy. Invest Ophthalmol Vis Sci 2001;42:1208-1214.

13.Howell N, Xu M, Halvorson S, Bodis-Wollner I, Sherman J. A heteroplasmic LHON family: tissue distribution and transmission of the 11778 mutation. Am J Hum Genet 1994;55:203-206.

14.Yen MY, Lee HC, Wang AG, Chang WL, Liu JH, Wei YH. Exclusive homoplasmic 11778 mutation in mitochondrial DNA of Chinese patients with Leber's hereditary optic neuropathy. Jpn J Ophthalmol 1999;43:196-200.

15.Chinnery PF, Howell N, Andrew RM, Turnbull DM. Mitochondrial DNA analysis: polymorphisms and pathogenicity. J Med Genet 1999;36:505-510.

16.Howell N, Mackey DA. Low-penetrance branches in matrilineal pedigrees with Leber hereditary optic neuropathy. Am J Hum Genet 1998;63:1220-1224.

17.Brown MD, Sun F, Wallace DC. Clustering of Caucasian Leber hereditary optic neuropathy patients containing the 11778 or 14484 mutations on an mtDNA lineage. Am J Hum Genet 1997;60:381-387.

18.Man PYW, Howell N, Mackey DA, Norby S, Rosenberg T, Trunbull DM, Chinnery PF. Mitochondrial DNA haplogroup distribution with Leber hereditary optic neuropathy pedigrees. J Med Genet 2004;41:e41.

19.Carelli V, Achilli A, Valentino ML, Rengo C, Semino O, Pala M, Olivieri A, Mattiazzi M, Pallotti F, Carrara F, Zeriani M, Leuzzi V, Carducci C, Valle G, Simionati B, Mendieta L, Salomao S, Belfort Jr R, Sadun AA, Torroni A. Haplogroup defects and recombination of mitochondrial DNA: novel clues from the analysis of Leber hereditary optic neuropathy pedigrees. Am J Hum Genet 2006;78:564-574.

20.Bu X, Rotter JI. X chromosome-linked and mitochondrial gene control of Leber hereditary optic neuropathy: evidence from segregation analysis for dependence on X-chromsome inactivation. Proc Natl Acad Sci USA 1991;88:8198-8202.

21.Hudson G, Keers S, Man PYW, Griffiths P, Huoponen K, Savontaus ML, Nikosckelainen E, Zeviani M, Carrara F, Horvath R, Karcagi V, Spruiji L, de Coo IFM, Smeets HJM, Chinnery PF. Identification of an X-chromosomal locus and haplotype modulating the phenotype of a mitochondrial DNA disorder. Am J Hum Genet 2005;77:1086-1091.

22.Shankar SP, Fingert JH, Carelli V, Valentino ML, King TM, Daiger SP, Salomao SR, Berezovsky A, Belfort Jr. R, Braun TA, Sheffield VC, Sadun AA, Stone EM. Evidence for a novel X-linked modifier locus for Leber hereditary optic neuropathy. Ophthalmic Genet 2008; 29: 17–24.

23.Abu-Amero KK, Jaber M, Hellani A, Bosley TM. Genome-wide expression profile of LHON patients with the 11778 mutation. Brit J Ophthalmol 2010:94:256-259.

24.Sadun AA, Carelli V, Salomao SR, Berezovsky A, Quiros PA, Sadun F, Denegri AM, Andrade R, Moraes M, Passos A, Kjaer P, Pereira J, Valentino ML, Schein S, Belfort R. Extensive investigation of a large Brazilian pedigree of 11778/haplogroup J Leber hereditary optic neuropathy. Am J Ophthalmol2003;136:231-238.

25.Kerrison JB, Miller NR, Hsu FC, Beaty TH, Maumenee IH, Smith KH, Savino PJ, Stone EM, Newman NJ. A case-control study of tobacco and alcohol consumption in Leber hereditary optic neuropathy. Am J Ophthalmol 2000;130:803-812.

26.Couse, J. F. and Korach, K. S. (1999) Estrogen receptor null mice: what have we learned and where will they lead us? *Endocr Rev* 20, 358-417.

27.Pettersson, K. and Gustafsson, J. A. (2001) Role of estrogen receptor β in estrogen action Annu Rev Physiol 63, 165-92.

28. Henderson, B. E., Ross, R. and Bernstein, L. (1988) Estrogens as a cause of human cancer: the Richard and Hinda Rosenthal Foundation award lecture Cancer Res 48, 246-53.

29. Konrad F. Koehler, Luisa A. et al. Reflections on the Discovery and Significance of Estrogen Receptor beta. Endocrine Reviews 2005;26(3):465–478.

30. de Voogd S, Wolfs RC, Jansonius NM, Uitterlinden AG, Pols HA, Hofman A, de Jong PT. Estrogen receptors alpha and beta and the risk of open-angle glaucoma: the Rotterdam Study. Arch Ophthalmol. 2008;126(1):110-4.

31. Carmeci C, Thompson DA, Ring HZ, Francke U, Weigel RJ. Identification of a gene (GPR30) with homology to the G-protein-coupled receptor superfamily associated with estrogen receptor expression in breast cancer. Genomics

1997;45:607-17.

32. Filardo EJ, Thomas P. GPR30: a seven-transmembrane-spanning estrogen receptor that triggers EGF release. Trends Endocrinol Metab 2005;16:362–7.

33. Brann DW, Dhandapani K, Wakade C, Mahesh VB, Khan MM. Neurotrophic and neuroprotective actions of estrogen: basic mechanisms and clinical implications. Steroids. 2007;72(5):381-405.

34. Giddabasappa A, Bauler M, Yepuru M, Chaum E, Dalton JT, Eswaraka J. 17- β estradiol protects ARPE-19 cells from oxidative stress through estrogen receptor- β . Invest Ophthalmol Vis Sci. 2010;51(10):5278-87.

35. Nonaka A, Kiryu J, Tsujikawa A, Yamashiro K, Miyamoto K, Nishiwaki H, Mandai M, Honda Y, Ogura Y. Administration of 17b-Estradiol Attenuates Retinal Ischemia–Reperfusion Injury in Rats. Invest Ophthalmol Vis Sci. 2000;41:2689–2696.

36. Kumar DM, Perez E, Cai ZY, Aoun P, Brun-ZinkernagelAM, Covey DF, Simpkins JW, Agarwal N. Role of nonfeminizing estrogen analogues in neuroprotection of rat retinal ganglion cells against glutamate-induced cytotoxicity. Free Radical Biol Med 2005;38: 1152–1163.

37. Kitaoka Y, Munemasa Y, Hayashi Y, Kuribayashi J, Koseki N, Kojima K, Kumai T, Ueno S. Axonal protection by 17β -estradiol through thioredoxin-1 in tumor necrosis factor-induced optic neuropathy. Endocrinology. 2011;152(7):2775-85.

38. Giordano C, Montopoli M, Perli E, Orlandi M, Fantin M, Ross-Cisneros FN, Caparrotta L, Martinuzzi A, Ragazzi E, Ghelli A, Sadun AA, d'Amati G, Carelli V. Oestrogens ameliorate mitochondrial dysfunction in Leber's hereditary optic neuropathy. Brain 2011;134:220-234.

39. Villena JA, Kralli A. ERRa: a metabolic function for the oldest orphan. Trends in Endocrinology and Metabolism 2008;19:269-276.

40.Hong H, Yang L, Stallcup MR. Hormone-independent transcriptional activation and coactivator binding by novel orphan nuclear receptor ERR3. J Biol Chem 1999;274:22618 –22626.

41.Giguere V, Yang N, Segui P, Evans RM. Identification of a new class of steroid hormone receptors. Nature 1988;331:91–94

42.Liu D, Zhang Z, Gladwell W, Teng CT. Estrogen stimulates estrogen-related receptor alpha gene expression through conserved hormone response elements. Endocrinology 2003;144(11):4894–4904.

43.Yang N, Shigeta H, Shi H, Teng CT. Estrogen-related receptor, hERR1, modulates estrogen receptor-mediated response of human lactoferrin gene promoter. J Biol Chem 1996;271:5795–5804

44.Zhang Z, Teng CT. Estrogen receptor-related receptor alpha 1 interacts with coactivator and constitutively activates the estrogen response elements of the human lactoferrin gene. J Biol Chem 2000;275:20837–20846

45.Zhang Z, Teng CT. Estrogen receptor alpha and estrogen receptor-related receptor alpha 1 compete for binding and coactivator. Mol Cell Endocrinol 2001;172: 223–233

46. Herzig, S., Long, F., Jhala, U. S., Hedrick, S., Quinn, R., Bauer, A., et al. CREB regulates hepatic gluconeogenesis through the coactivator PGC-1. *Nature* 2001;413, 179–183.

47. Yoon, J. C., Puigserver, P., Chen, G., Donovan, J., Wu, Z., Rhee, J., et al. Control of hepatic gluconeogenesis through the transcriptional coactivator PGC-1. *Nature* 2001; 413, 131–138.

48.Kincaid B., Ella Bossy-Wetzel E. Forever young: SIRT3 a shield against mitochondrial meltdown, aging, and neurodegeneration. Frontiers in Aging Neuroscience 2013; 5:1-13.

49. Kong, X., Wang, R., Xue, Y., Liu, X., Zhang, H., Chen, Y., et al.. Sirtuin 3, a new target of

PGC-1alpha, plays an important role in the suppression of ROS and mitochondrial biogenesis. *PLoS ONE* 2010; 5:e11707.

50.Pillai, V. B., Sundaresan, N. R., Kim, G., Gupta, M., Rajamohan, S. B., Pillai, J. B., et al. Exogenous NAD blocks cardiac hypertrophic response via activation of the SIRT3-LKB1-AMP-activated kinase pathway. *J. Biol. Chem.* 2010; 285, 3133–3144.

51.Woods, A., Johnstone, S. R., Dickerson, K., Leiper, F. C., Fryer, L. G. D., Neumann, D., et al. LKB1 is the upstream kinase in the AMP-activated protein kinase cascade. *Curr. Biol.* 2003;13, 2004–2008.

52.Bergeron, R., Ren, J. M., Cadman, K. S., Moore, I. K., Perret, P., Pypaert, M., et al. Chronic activation of AMP kinase results in NRF-1 activation and mitochondrial biogenesis. *Am. J. Physiol. Endocrinol. Metab.* 2001; 281, E1340–E1346.

53. Lee S, Van Bergen NJ, Kong GY et al. Mitochondrial dysfunction in glaucoma and emerging bioenergetic therapies. Experimental Eye Research 93 (2011) 204-212.

54. Man PY, Griffiths PG, Brown DT, Howell N, Turnbull DM, Chinnery PF. The epidemiology of Leber hereditary optic neuropathy in the North East of England. Am J Hum Genet. 2003;72(2):333-9.

五. 計畫成果自評

Leber's hereditary optic neuropathy (LHON) is characterized by acute and subacute visual loss predominantly affecting young man. It is a maternally transmitted disease caused by mitochondria DNA (mtDNA) mutation, which is transmitted to all the maternal lineages. However, there is an unexplained male prevalence in LHON that over 80% of LHON patients are male. These low penetrance and male prevalence may result from undefined neuroprotective mechanism which may be beneficial for future treatment. We propose this project to explore the role of estrogen-related receptors (ERR α , ERR β and ERR γ) in the pathogenesis of LHON especially focusing on gender difference.

We have examined the expression of estrogen-related receptors (ERR α , ERR β and ERR γ) in mouse retina. ERR α and ERR γ proteins are both widely expressed over the whole retinal layers, with intense level over ganglion cell layer and photoreceptor inner segment. ERR β is expressed mainly over inner retina, with the outer retina relatively spared. Its expression corresponds to the span of Muller cell processes, and was co-localized with Muller cell marker glutamine synthetase.

Following optic nerve injury, we observe the expressional change of estrogen-related receptors (ERR α , ERR β and ERR γ) in the injured retina. These three estrogen-related receptors showed a similar trend that their expression in the ganglion cell layer increased gradually and peaked around 3 days after injury, and then declined at 5 days and 7 days thereafter. We did not observe gender difference of estrogen-related receptors expression throughout the whole course. Western blot analysis showed a similar pattern of expressional change after optic nerve injury, which may reflect neuronal cell response to axonal injury.

科技部補助計畫衍生研發成果推廣資料表

日期:2015/10/16

	計畫名稱:利伯氏遺傳視神經病變之雌激素相關受器與性別差異研究						
科技部補助計畫	計畫主持人:王安國						
	計畫編號: 103-2629-B-075-001-	學門領域: 性別主流科技計畫					
	無研發成果推廣	資料					

103年度專題研究計畫研究成果彙整表

	1007 及于赵州九时重州九风小禾正代
計畫主持人: 王安國	計畫編號:103-2629-B-075-001-
計畫夕稱 :利伯氏 溃值 進	抽纸店繼之雌激去扣閯恶哭的财别关胃研究

計畫名稱: 利伯氏遺傳視神經病變之雌激素相關受器與性別差異研究							
成果項目			量化				備註(質化說明
			數(被接受	預期總達成 數(含實際 已達成數)		單位	:如數個計畫共 同成果、成果列 為該期刊之封面 故事等)
		期刊論文	1	0	0%		論文撰稿中
	論文著作	研究報告/技術報告	0	0	100%	篇	
		研討會論文	1	0	0%		論文撰稿中
		專書	0	0	100%	章/本	
	声 エル	申請中件數	0	0	100%	14	
围内	專利	已獲得件數	0	0	100%	件	
國內	计化放神	件數	0	0	100%	件	
	技術移轉	權利金	0	0	100%	千元	
		碩士生	0	0	100%		
	參與計畫人力	博士生	0	0	100%	人次	
	(本國籍)	博士後研究員	0	0	100%		
		專任助理	1	1	100%		
	み 上 枯 ル	期刊論文	1	0	0%	篇	論文撰稿中
		研究報告/技術報告	0	0	100%		
	論文著作	研討會論文	1	0	0%		論文撰稿中
		專書	0	0	100%	章/本	
	声 い	申請中件數	0	0	100%	件	
國外	專利	已獲得件數	0	0	100%	什	
國外	计你的抽	件數	0	0	100%	件	
	技術移轉	權利金	0	0	100%	千元	
	參與計畫人力 (外國籍)	碩士生	0	0	100%		
		博士生	0	0	100%	Leb	
		博士後研究員	0	0	100%	人次	
		專任助理	0	0	100%		
成、際際產效	其以辨項 法如得作響技事述 是以辨獎、力術項 是 之 動國國助 體 文 、	已完成動物實驗, 器(ERRα, ERRβ a 術(optic nerve cr 變化,此外,特別著	nd ERRγ), ush),檢查,	在小鼠視網 1 1 鼠視網膜在	莫的表現, 受傷後之:	並利用; 三種雌激	視神經壓迫傷手 o素相關受器表現

	成果項目	量化	名稱或內容性質簡述
	測驗工具(含質性與量性)	0	
科 教	課程/模組	0	
一處	電腦及網路系統或工具	0	
計 -	教材	0	
畫加	舉辦之活動/競賽	0	
填	研討會/工作坊	0	
項 目	電子報、網站	0	
	計畫成果推廣之參與(閱聽)人數	0	

科技部補助專題研究計畫成果報告自評表

請就研究內容與原計畫相符程度、達成預期目標情況、研究成果之學術或應用價值(簡要敘述成果所代表之意義、價值、影響或進一步發展之可能性)、是否適 合在學術期刊發表或申請專利、主要發現或其他有關價值等,作一綜合評估。

1.	請就研究內容與原計畫相符程度、達成預期目標情況作一綜合評估 ■達成目標 □未達成目標(請說明,以100字為限) □實驗失敗 □因故實驗中斷 □其他原因
	說明:
2.	研究成果在學術期刊發表或申請專利等情形: 論文: \Box 已發表 \Box 未發表之文稿 ■ 撰寫中 \Box 無 專利: \Box 已獲得 \Box 申請中 ■ 無 技轉: \Box 已技轉 \Box 洽談中 ■ 無 其他: (以100字為限) 已完成動物實驗,利用螢光免疫染色與西方墨點試驗,檢查三種雌激素相關受 器(ERR α , ERR β and ERR γ),在小鼠視網膜的表現,並利用視神經壓迫傷手 術(optic nerve crush),檢查小鼠視網膜在受傷後之三種雌激素相關受器表 現變化,此外,特別著重於雄鼠與雌鼠的表現差異性,我們的論文在撰稿中。
3.	請依學術成就、技術創新、社會影響等方面,評估研究成果之學術或應用價值 (簡要敘述成果所代表之意義、價值、影響或進一步發展之可能性)(以 500字為限) 利伯氏遺傳視神經病變(LHON)是一種母系遺傳的粒線體突變,它會造成急性視 力減退,好發於男性,佔80%,視力常惡化至零點一以下,伴隨視野缺損,兩 眼可同時發病。粒線體突變會傳給所有母系細胞,然而具有突變基因的男性發 病率僅有50%,女性更只有10%,顯示其發病率低與好發於男性的特點。 其發病率低與好發於男性的特點,目前無明確原因,推測可能源於神經保 護機轉,若能瞭解此機轉可能有助於未來的治療,雌激素與其衍生物已証明具 有神經保護作用,我們提出此計畫來探討雌激素相關受器(Estrogen-related receptor, ERRs)在視神經受損所扮演的角色,特別著重於其性別差異,若能 瞭解此機轉可能有助於未來的治療,。